

Package ‘PwrGSD’

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Title Power in a Group Sequential Design

Version 1.0

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Depends survival

Description Tools to compute power in a group sequential design. SimPwrGSD C-kernel is a simulation routine that is similar in spirit to dssp2.f by Gu and Lai, but with major improvements. AsyPwrGSD has exactly the same range of application as SimPwrGSD but uses asymptotic methods and runs much faster.

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R topics documented:

PwrGSD	1
cpd.PwrGSD	9
Power	13
Elements	14
plot.cpd.PwrGSD	14
GrpSeqBnds	15
SimGSB	19
as.boundaries	20
wtdlogrank	21
IntSurvDiff	22
mysurvfit	24
agghaz	25
mystack	26
CDFOR2LRR	27
CY2TOShaz	28
CRRtoRR	28
RCM2RR	29
RR2RCM	30
lookup	31
lung	31
Surv	32
DX	33
paste	34

Description

Derives power in a two arm clinical trial under a group sequential design. Allows for arbitrary number of interim analyses, arbitrary specification of arm-0/arm-1 time to event distributions (via survival or hazard), arm-0/arm-1 censoring distribution, provisions for two types of continuous time non-compliance according to arm-0/arm-1 rate followed by switch to new hazard rate. Allows for analyses using (I) weighted log-rank statistic, with weighting function (a) a member of the Fleming-Harrington G-Rho class, or (b) a stopped version thereof, or (c) the ramp-plateau deterministic weights, or (II) the integrated survival distance (currently under method=="S" without futility only). Stopping boundaries are computed via the Lan-Demets method, Haybittle method, or converted from the stochastic curtailment procedure. The Lan-Demets boundaries can be constructed using either O'Brien-Fleming, Pocock or Wang-Tsiatis power alpha-spending. The C kernel is readily extensible, and further options will become available in the near future.

Usage

```
PwrGSD(EfficacyBoundary = LanDemets(alpha = 0.05, spending = ObrienFleming),
       FutilityBoundary = LanDemets(alpha = 0.1, spending = ObrienFleming),
       sided = c("2", ">", "<"), method = c("S", "A"), accrue, accrat,
       tlook, tcut0 = NULL, h0 = NULL, s0 = NULL, tcut1 = NULL,
       rhaz = NULL, h1 = NULL, s1 = NULL, tcutc0 = NULL, hc0 = NULL,
       sc0 = NULL, tcutc1 = NULL, hc1 = NULL, sc1 = NULL, tcutd0A = NULL,
       hd0A = NULL, sd0A = NULL, tcutd0B = NULL, hd0B = NULL, sd0B = NULL,
       tcutd1A = NULL, hd1A = NULL, sd1A = NULL, tcutd1B = NULL,
       hd1B = NULL, sd1B = NULL, tcutx0A = NULL, hx0A = NULL, sx0A = NULL,
       tcutx0B = NULL, hx0B = NULL, sx0B = NULL, tcutx1A = NULL,
       hx1A = NULL, sx1A = NULL, tcutx1B = NULL, hx1B = NULL, sx1B = NULL,
       noncompliance = c("none", "crossover", "mixed", "user"),
       gradual = FALSE, WtFun = c("FH", "SFH", "Ramp"), ppar = cbind(c(0, 0)),
       Spend.Info = c("Variance", "Events", "Hybrid(k)", "Calendar"), RR.Futility = NULL,
       qProp.one.or.Q = c("one", "Q"), Nsim = NULL, detail = FALSE, StatType = c("WLR",
       "ISD"))
```

Arguments

EfficacyBoundary

This specifies the method used to construct the efficacy boundary. The available choices are

- (i) `Lan-Demets(alpha=<total type I error>, spending=<spending function>)`. The Lan-Demets method is based upon a error probability spending approach. The spending function can be set to `ObrienFleming`, `Pocock`, or `Power(rho)`, where `rho` is the the power argument for the power spending function: `rho=3` is roughly equivalent to the O'Brien-Fleming spending function and smaller powers result in a less conservative spending function.
- (ii) `Haybittle(alpha=<total type I error>, b.Haybittle=<user specified boundary point>)`. The Haybittle approach is the simplest, which sets

the boundary points equal to `b.Haybittle`, a user specified value (try 3) for all analyses except the last, which is calculated so as to result in the total type I error, set with the argument `alpha`.

- (iii) `SC(alpha=<total type I error>, crit=<threshold for conditional type I error for efficacy stopping>)`. The stochastic curtailment method is based upon the conditional probability of type I error given the current value of the statistic. Under this method, a sequence of boundary points on the standard normal scale (as are boundary points under all other methods) is calculated so that the total probability of type I error, `alpha`, is maintained. This is done by considering the joint probabilities of continuing to the current analysis and then exceeding the threshold at the current analysis. A good value for the threshold value for the conditional type I error, `crit` is 0.90 or greater.

- (iv) User supplied boundary points in the form `c(b1, b2, b3, ..., b_m)`, where `m` is the number of looks.

FutilityBoundary

This specifies the method used to construct the futility boundary. The available choices are

- (i) `Lan-Demets(alpha=<total type II error>, spending=<spending function>)`. The Lan-Demets method is based upon a error probability spending approach. The spending function can be set to `ObrienFleming`, `Pocock`, or `Power(rho)`, where `rho` is the the power argument for the power spending function: `rho=3` is roughly equivalent to the O'Brien-Fleming spending function and smaller powers result in a less conservative spending function.
- (ii) `Haybittle(alpha=<total type I error>, b.Haybittle=<user specified boundary point>)`. The Haybittle approach is the simplest, which sets the boundary points equal to `b.Haybittle`, a user specified value (try 3) for all analyses except the last, which is calculated so as to result in the total type II error, set with the argument `alpha`.
- (iii) `SC(alpha=<total type II error>, crit=<threshold for conditional type II error for futility stopping>, drift.end=<projected drift at end of trial>)`. The stochastic curtailment method is based upon the conditional probability of type II error given the current value of the statistic. Under this method, a sequence of boundary points on the standard normal scale (as are boundary points under all other methods) is calculated so that the total probability of type II error, `alpha`, is maintained. This is done by considering the joint probabilities of continuing to the current analysis and then exceeding the threshold at the current analysis. A good value for the threshold value for the conditional type I error, `crit` is 0.90 or greater.
- (iv) User supplied boundary points in the form `c(b1, b2, b3, ..., b_m)`, where `m` is the number of looks.

sided If two-sided tests of H_0 , set to "2" (quoted). If one-sided test of H_0 , set to ">" for upper tail, "<" for lower tail. If `method=="S"` then this must be of the same length as `StatType` because the interpretation of `sided` is different depending upon whether `StatType=="WLR"` (negative is benefit) or `StatType=="ISD"` (positive is benefit)

method Determines how to calculate the power. Set to "A" (Asymptotic method, the default) or "S" (Simulation method)

<code>accru</code>	The upper endpoint of the accrual period beginning with time 0.
<code>accrat</code>	The rate of accrual per unit of time.
<code>tlook</code>	The times of planned interim analyses.
<code>tcut0</code>	Left hand endpoints for intervals upon which the arm-0 specific mortality is constant. The last given component is the left hand endpoint of the interval having right hand endpoint infinity.
<code>h0</code>	A vector of the same length as <code>tcut0</code> which specifies the piecewise constant arm-0 mortality rate.
<code>s0</code>	Alternatively, the arm-0 mortality distribution can be supplied via this argument, in terms of the corresponding survival function values at the times given in the vector <code>tcut0</code> . If <code>s0</code> is supplied, then <code>h0</code> is derived internally, assuming the piecewise exponential distribution. If you specify <code>s0</code> , the first element must be 1, and correspondingly, the first component of <code>tcut0</code> will be the lower support point of the distribution. You must supply either <code>h0</code> or <code>s0</code> but not both.
<code>tcut1</code>	Left hand endpoints for intervals upon which the arm-1 specific mortality is constant. The last given component is the left hand endpoint of the interval having right hand endpoint infinity.
<code>rhaz</code>	A vector of piecewise constant arm-1 versus arm-0 mortality rate ratios. If <code>tcut1</code> and <code>tcut0</code> are not identical, then <code>tcut1</code> , <code>h0</code> , and <code>rhaz</code> are internally rederived at the union of the sequences <code>tcut0</code> and <code>tcut1</code> . In all cases the arm-1 mortality rate is then derived at the time cutpoints <code>tcut1</code> as <code>rhaz</code> \times <code>h0</code> .
<code>h1</code>	Alternatively, the arm-1 mortality distribution can be supplied via this argument by specifying the piecewise constant arm-1 mortality rate. See the comments above.
<code>s1</code>	Alternatively, the arm-1 mortality distribution can be supplied via this argument, in terms of the corresponding survival function values at the times given in the vector <code>tcut1</code> . Comments regarding <code>s0</code> above apply here as well. You must supply exactly one of the following: <code>h1</code> , <code>rhaz</code> , or <code>s1</code> .
<code>tcutc0</code>	Left hand endpoints for intervals upon which the arm-0 specific censoring distribution hazard function is constant. The last given component is the left hand endpoint of the interval having right hand endpoint infinity.
<code>hc0</code>	A vector of the same length as <code>tcutc0</code> which specifies the arm-0 censoring distribution in terms of a piecewise constant hazard function.
<code>sc0</code>	Alternatively, the arm-0 censoring distribution can be supplied via this argument, in terms of the corresponding survival function values at the times given in the vector <code>tcutc0</code> . See comments above. You must supply either <code>hc0</code> or <code>sc0</code> but not both.
<code>tcutc1</code>	Left hand endpoints for intervals upon which the arm-1 specific censoring distribution hazard function is constant. The last given component is the left hand endpoint of the interval having right hand endpoint infinity.
<code>hc1</code>	A vector of the same length as <code>tcutc1</code> which specifies the arm-1 censoring distribution in terms of a piecewise constant hazard function.
<code>sc1</code>	Alternatively, the arm-1 censoring distribution can be supplied via this argument, in terms of the corresponding survival function values at the times given in the vector <code>tcutc1</code> . See comments above. You must supply either <code>hc1</code> or <code>sc1</code> but not both.

noncompliance	(i) Setting noncompliance to “none” for no non-compliance will automatically set the non-compliance arguments, below, to appropriate values for no compliance. This requires no additional user specification of non-compliance parameters. (ii) Setting noncompliance to “crossover” will automatically set crossover values in the arm 0/1 specific <i>post-cause-B-delay-mortality</i> for cross-over, i.e. simple interchange of the arm 0 and arm 1 mortalities. The user is required to specify all parameters corresponding to the arm 0/1 specific <i>cause-B-delay</i> distributions. The <i>cause-A-delay</i> and <i>post-cause-A-delay-mortality</i> are automatically set so as not to influence the calculations. Setting noncompliance to “mixed” will set the arm 0/1 specific <i>post-cause-B-delay-mortality</i> distributions for crossover as defined above. The user specifies the arm 0/1 specific <i>cause-B-delay</i> distribution as above, and in addition, all parameters related to the arm 0/1 specific <i>cause-A-delay</i> distributions and corresponding arm 0/1 specific <i>post-cause-A-delay-mortality</i> distributions. (iii) Setting noncompliance to “user” requires the user to specify all non-compliance distribution parameters.
tcutd0A	Left hand endpoints for intervals upon which the arm-0 specific <i>cause-A delay</i> distribution hazard function is constant. The last given component is the left hand endpoint of the interval having right hand endpoint infinity. Required only when noncompliance is set to “mixed” or “user”.
hd0A	A vector of the same length as tcutd0A containing peicewise constant hazard rates for the arm-0 <i>cause-A delay</i> distribution. Required only when noncompliance is set to “mixed” or “user”.
sd0A	When required, the arm-0 <i>cause-A-delay</i> distribution is alternately specified via a survival function. A vector of the same length as tcutd0A .
tcutd0B	Left hand endpoints for intervals upon which the arm-0 specific <i>cause-B delay</i> distribution hazard function is constant. The last given component is the left hand endpoint of the interval having right hand endpoint infinity. Always required when noncompliance is set to any value other than “none”.
hd0B	A vector of the same length as tcutd0B containing peicewise constant hazard rates for the arm-0 <i>cause-B delay</i> distribution. Always required when noncompliance is set to any value other than “none”.
sd0B	When required, the arm-0 <i>cause-B-delay</i> distribution is alternately specified via a survival function. A vector of the same length as tcutd0B .
tcutd1A	Left hand endpoints for intervals upon which the arm-1 specific <i>cause-A delay</i> distribution hazard function is constant. The last given component is the left hand endpoint of the interval having right hand endpoint infinity. Required only when noncompliance is set to “mixed” or “user”.
hd1A	A vector of the same length as tcutd1A containing peicewise constant hazard rates for the arm-1 <i>cause-A delay</i> distribution. Required only when noncompliance is set to “mixed” or “user”.
sd1A	When required, the arm-1 <i>cause-A-delay</i> distribution is alternately specified via a survival function. A vector of the same length as tcutd1A .
tcutd1B	Left hand endpoints for intervals upon which the arm-1 specific <i>cause-B delay</i> distribution hazard function is constant. The last given component is the left hand endpoint of the interval having right hand endpoint infinity. Always required when noncompliance is set to any value other than “none”.

hd1B	A vector of the same length as <code>tcutd1B</code> containing peicewise constant hazard rates for the arm-1 <i>cause-B delay</i> distribution. Always required when <code>noncompliance</code> is set to any value other than “none”.
sd1B	When required, the arm-1 <i>cause-A-delay</i> distribution is alternately specified via a survival function. A vector of the same length as <code>tcutd1A</code> .
tcutx0A	Left hand endpoints for intervals upon which the arm-0 specific <i>post-cause-A-delay-mortality</i> rate is constant. The last given component is the left hand endpoint of the interval having right hand endpoint infinity. Required only when <code>noncompliance</code> is set to “mixed” or “user”.
hx0A	A vector of the same length as <code>tcutx0A</code> containing the arm-0 <i>post-cause-A-delay mortality</i> rates. Required only when <code>noncompliance</code> is set to “mixed” or “user”.
sx0A	When required, the arm-0 <i>post-cause-A-delay mortality</i> distribution is alternately specified via a survival function. A vector of the same length as <code>tcutx0A</code> .
tcutx0B	Left hand endpoints for intervals upon which the arm-0 specific <i>post-cause-B-delay-mortality</i> rate is constant. The last given component is the left hand endpoint of the interval having right hand endpoint infinity. Always required when <code>noncompliance</code> is set to any value other than “none”.
hx0B	A vector of the same length as <code>tcutx0B</code> containing the arm-0 <i>post-cause-B-delay mortality</i> rates. Always required when <code>noncompliance</code> is set to any value other than “none”.
sx0B	When required, the arm-0 <i>post-cause-B-delay mortality</i> distribution is alternately specified via a survival function. A vector of the same length as <code>tcutx0B</code> .
tcutx1A	Left hand endpoints for intervals upon which the arm-1 specific <i>post-cause-A-delay-mortality</i> rate is constant. The last given component is the left hand endpoint of the interval having right hand endpoint infinity. Required only when <code>noncompliance</code> is set to “mixed” or “user”.
hx1A	A vector of the same length as <code>tcutx1A</code> containing the arm-1 <i>post-cause-A-delay mortality</i> rates. Required only when <code>noncompliance</code> is set to “mixed” or “user”.
sx1A	When required, the arm-1 <i>post-cause-A-delay mortality</i> distribution is alternately specified via a survival function. A vector of the same length as <code>tcutx1A</code> .
tcutx1B	Left hand endpoints for intervals upon which the arm-1 specific <i>post-cause-B-delay-mortality</i> rate is constant. The last given component is the left hand endpoint of the interval having right hand endpoint infinity. Always required when <code>noncompliance</code> is set to any value other than “none”.
hx1B	A vector of the same length as <code>tcutx1B</code> containing the arm-1 <i>post-cause-B-delay mortality</i> rates. Always required when <code>noncompliance</code> is set to any value other than “none”.
sx1B	When required, the arm-1 <i>post-cause-B-delay mortality</i> distribution is alternately specified via a survival function. A vector of the same length as <code>tcutx1B</code> .
gradual	Should the conversion to post-noncompliance mortality be gradual. Under the default behavior, <code>gradual=FALSE</code> , there is an immediate conversion to the post-noncompliance mortality rate function. If <code>gradual</code> is set to

	TRUE then this conversion is done “gradually”. In truth, at the individual level what is done is that the new mortality rate function is a convex combination of the pre-noncompliance mortality and the post-noncompliance mortality, with the weighting in proportion to the time spent in compliance with the study arm protocol.
WtFun	Specifies the name of a weighting function (of time) for assigning relative weights to events according to the times at which they occur. The default, “FH”, a two parameter weight function, specifies the ‘Fleming-Harrington’ g - ρ family of weighting functions defined as the pooled arm survival function (Kaplan-Meier estimate) raised to the g times its complement raised to the ρ . Note that $g=\rho=0$ corresponds to the unweighted log-rank statistic. A second choice is the “SFH” function, (for ‘Stopped Fleming-Harrington’), meaning that the “FH” weights are capped at their value at a user specified time, which has a total of 3 parameters. A third choice is <code>Ramp(tcutoff)</code> . Under this choice, weights are assigned in a linearly manner from time 0 until a user specified cut-off time, <code>tcutoff</code> , after which events are weighted equally. It is possible to conduct computations on <code>nstat</code> candidate statistics within a single run. In this case, <code>WtFun</code> should be a character vector of length <code>nstat</code> having components set from among the available choices.
ppar	A vector containing all the weight function parameters, in the order determined by that of “WtFun”. For example, if <code>WtFun</code> is set to <code>c("FH", "SFH", "Ramp")</code> then <code>ppar</code> should be a vector of length six, with the “FH” parameters in the first two elements, “SFH” parameters in the next 3 elements, and “Ramp” parameter in the last element.
RR.Futility	The relative risk corresponding to the alternative hypothesis that is required in the construction of the futility boundary. Required if <code>Boundary.Futility</code> is set to a non-null value.
Spend.Info	When the test statistic is something other than the unweighted log-rank statistic, the variance information, i.e. the ratio of variance at interim analysis to variance at the end of trial, is something other than the ratio of events at interim analysis to the events at trial end. The problem is that in practice one doesn’t necessarily have a good idea what the end of trial variance should be. In this case the user may wish to spend the type I and type II error probabilities according to a different time scale. Possible choices are “Variance”, (default), which just uses the variance ratio scale, “Events”, which uses the events ratio scale, “Hybrid(k)”, which makes a linear transition from the “Variance” scale to the “Events” scale beginning with analysis number k . The last choice, “Calendar”, uses the calendar time scale
qProp.one.or.Q	If a futility boundary is specified, what assumption should be made about the drift function (the mean value of the weighted log-rank statistic at analysis j normalized by the square root of the variance function at analysis k). In practice we don’t presume to know the shape of the drift function. Set to “one” or “Q”. The choice “one” results in a more conservative boundary.
Nsim	If you specify <code>method=="S"</code> , then you must specify the number of simulations. 1000 should be sufficient.
detail	If you specify <code>method=="S"</code> , and want to see the full level of detail regarding arguments returned from the C level code, specify <code>detail==TRUE</code>

StatType If you specify `method=="S"`, then the available choices are "WLR" (weighted log-rank) and "ISD" (integrated survival difference).

Value

Returns a value of class **PwrGSD** which has components listed below. Note that the `print` method will display a summary table of estimated powers and type I errors as a `nstat` by 2 matrix. The summary method returns the same object invisibly, but after computing the summary table mentioned above, and it is included in the returned value as a component `TBL`. See examples below.

dPower	A <code>length(tlook)</code> by <code>nstat</code> matrix containing in each column, an increment in power that resulted at that analysis time for the given statistic.
dErrorI	A <code>length(tlook)</code> by <code>nstat</code> matrix containing in each column, an increment in type I error that resulted at that analysis time for the given statistic. Always sums to the total alpha specified in <code>alphatot</code>
detail	A list with components equal to the arguments of the C-call, which correspond in a natural way to the arguments specified in the R call, along with the computed results in <code>palpha0vec</code> , <code>palpha1vec</code> , <code>inffrac</code> , and <code>mu</code> . The first two are identical to <code>dErrorI</code> and <code>dPower</code> , explained above. The last two are <code>length(tlook)</code> by <code>nstat</code> matrices. For each statistic specified in <code>par</code> , the corresponding columns of <code>pinffrac</code> and <code>mu</code> contain the information fraction and drift at each of the analysis times.
call	the call

Author(s)

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References

- Gu, M.-G. and Lai, T.-L. "Determination of power and sample size in the design of clinical trials with failure-time endpoints and interim analyses." *Controlled Clinical Trials* 20 (5): 423-438. 1999
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- Jennison, C. and Turnbull, B.W. (1999) *Group Sequential Methods: Applications to Clinical Trials* Chapman & Hall/Crc, Boca Raton FL
- Proschan, M.A., Lan, K.K.G., Wittes, J.T. (2006), corr 2nd printing (2008) *Statistical Monitoring of Clinical Trials A Unified Approach* Springer Verlag, New York

See Also

`cpd.PwrGSD`

Examples

```
library(PwrGSD)

test.example <-
  PwrGSD(EfficacyBoundary = LanDemets(alpha = 0.05, spending = ObrienFleming),
        FutilityBoundary = LanDemets(alpha = 0.1, spending = ObrienFleming),
        RR.Futility = 0.82, sided="<", method="A", accru = 7.73, accrat = 9818.65,
```



```

tlook =c(7.14, 8.14, 9.14, 10.14, 10.64, 11.15, 12.14, 13.14,
         14.14, 15.14, 16.14, 17.14, 18.14, 19.14, 20.14),
tcut0 =0:19, h0 =c(rep(3.73e-04, 2), rep(7.45e-04, 3),
                  rep(1.49e-03, 15)),
tcut1 =0:19, rhaz =c(1, 0.9125, 0.8688, 0.7814, 0.6941,
                   0.6943, 0.6072, 0.5202, 0.4332, 0.6520,
                   0.6524, 0.6527, 0.6530, 0.6534, 0.6537,
                   0.6541, 0.6544, 0.6547, 0.6551, 0.6554),
tcutc0 =0:19, hc0 =c(rep(1.05e-02, 2), rep(2.09e-02, 3),
                   rep(4.19e-02, 15)),
tcutc1 =0:19, hc1 =c(rep(1.05e-02, 2), rep(2.09e-02, 3),
                   rep(4.19e-02, 15)),
tcutd0B =c(0, 13), hd0B =c(0.04777, 0),
tcutd1B =0:6, hd1B =c(0.1109, 0.1381, 0.1485, 0.1637, 0.2446,
                    0.2497, 0),
noncompliance =crossover, gradual =TRUE,
WtFun =c("FH", "SFH", "Ramp"),
ppar =c(0, 1, 0, 1, 10, 10))

```

cpd.PwrGSD

Create a skeleton compound PwrGSD object

Description

Given a user defined indexing dataframe as its only argument, creates a skeleton compound PwrGSD object having a component **Elements**, a list of PwrGSD objects, of length equal to the number of rows in the indexing dataframe

Usage

```
cpd.PwrGSD(descr)
```

Arguments

descr	A dataframe of a number of rows equal to the length of the resulting list, Elements , of PwrGSD objects. The user defines the mapping between rows of descr and components of Elements and uses it to set up a loop over scenarios. There are several S3 classes and methods for example <code>plot.cpd.PwrGSD</code> , which exploit this mapping between characteristics of a run and the rows of descr for subsetting and constructing conditioned plots. See the example below.
--------------	-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------

Value

An object of class `cpd.PwrGSD` containing elements:

date	the POSIX date that the object was created—its quite useful
Elements	a list of length equal to the number of rows of descr which will later contain objects of class PwrGSD
descr	a copy of the indexing dataframe argument for use in navigating the compound object in subsequent calls to other functions such as the related <code>plot</code> method, and the subset extractor, Elements

Note

A `cpd.PwrGSD` object essentially a list of `PwrGSD` objects that a user may set up in order to investigate the space of possible trial scenarios, test statistics, and boundary construction options. One could store a list of results without appealing at all to these internal indexing capabilities. The advantage of setting up a `cpd.PwrGSD` object is the nice summarization functionality provided, for example the `plot` method for the `cpd.PwrGSD` class.

The key ingredient to (i) the construction of the empty object, (ii) and summarizing the results in tabular or plotted form via its manipulation in subsequent function calls, is the indexing dataset, `descr` (for description). The correspondence between rows of `descr` and elements in the list of `PwrGSD` objects is purposely left very loose. In the example outlined below, the user creates a “base case” call to `PwrGSD` and then decides which quantities in this “base case” call to vary in order to navigate the space of possible trial scenarios, monitoring statistics and boundary construction methods. Next, for each one of these settings being varied, a variable with levels that determine each possible setting is created. The dataset `descr` is created with one line corresponding to each combination of the selection variables so created. In order to ensure that there is 1-1 correspondence between the order of the rows in `descr` and the order in the list `Elements` of `PwrGSD` objects, the user carries out the computation in a loop over rows of `descr` in which the values of the selection variables in each given row of `descr` are used to create the corresponding component of `Elements` via an update the “base case” call.

Author(s)

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See Also

`Elements`, `plot.cpd.PwrGSD` and `Power`

Examples

```
## don't worry--these examples are guaranteed to work,
## its just inconvenient for the package checker
## Not run:
library(PwrGSD)

## In order to set up a compound object of class `cpd.PwrGSD'
## we first construct a base case: a two arm trial randomized in just
## under eight years with a maximum of 20 years of follow-up.
## We compute power at a specific alternative, `rhaz', under
## an interim analysis plan with roughly one annual analysis, some
## crossover between intervention and control arms, with Efficacy
## and futility boundaries constructed via the Lan-Demets procedure
## with O'Brien-Fleming spending on the hybrid scale. Investigate
## the behavior of three weighted log-rank statistics.

test.example <-
  PwrGSD(EfficacyBoundary = LanDemets(alpha = 0.05, spending = ObrienFleming),
        FutilityBoundary = LanDemets(alpha = 0.1, spending = ObrienFleming),
        RR.Futility = 0.82, sided="<",method="A",accru =7.73, accrat =9818.65,
        tlook =c(7.14, 8.14, 9.14, 10.14, 10.64, 11.15, 12.14, 13.14,
                  14.14, 15.14, 16.14, 17.14, 18.14, 19.14, 20.14),
        tcut0 =0:19, h0 =c(rep(3.73e-04, 2), rep(7.45e-04, 3),
                           rep(1.49e-03, 15)),
```

```

tcut1 =0:19, rhaz =c(1, 0.9125, 0.8688, 0.7814, 0.6941,
                    0.6943, 0.6072, 0.5202, 0.4332, 0.6520,
                    0.6524, 0.6527, 0.6530, 0.6534, 0.6537,
                    0.6541, 0.6544, 0.6547, 0.6551, 0.6554),
tcutc0 =0:19, hc0 =c(rep(1.05e-02, 2), rep(2.09e-02, 3),
                    rep(4.19e-02, 15)),
tcutc1 =0:19, hc1 =c(rep(1.05e-02, 2), rep(2.09e-02, 3),
                    rep(4.19e-02, 15)),
tcutd0B =c(0, 13), hd0B =c(0.04777, 0),
tcutd1B =0:6, hd1B =c(0.1109, 0.1381, 0.1485, 0.1637, 0.2446,
                    0.2497, 0),
noncompliance =crossover, gradual =TRUE,
WtFun =c("FH", "SFH", "Ramp"),
ppar =c(0, 1, 0, 1, 10, 10))

## we will construct a variety of alternate hypotheses relative to the
## base case specified above

rhaz <-
  c(1, 0.9125, 0.8688, 0.7814, 0.6941, 0.6943, 0.6072, 0.5202, 0.4332,
    0.652, 0.6524, 0.6527, 0.653, 0.6534, 0.6537, 0.6541, 0.6544,
    0.6547, 0.6551, 0.6554)

max.effect <- 0.80 + 0.05*(0:8)
n.me <- length(max.effect)

## we will also vary extent of censoring relative to the base case
## specified above

hc <- c(rep(0.0105, 2), rep(0.0209, 3), rep(0.0419, 15))

cens.amt <- 0.75 + 0.25*(0:2)
n.ca <- length(cens.amt)

## we may also wish to compare the Lan-Demets/O'Brien-Fleming efficacy
## boundary with a Pocock efficacy boundary

Eff.bound.choice <- 1:2
ebc.nms <- c("LanDemets(alpha=0.05, spending=ObrienFleming)",
            "SC(alpha=0.05, crit=0.90)")
n.ec <- length(Eff.bound.choice)

## The following line creates the indexing dataframe, `descr', with one
## line for each possible combination of the selection variables we've
## created.

descr <- as.data.frame(
  cbind(Eff.bound.choice=rep(Eff.bound.choice, each=n.ca*n.me),
        cens.amt=rep(rep(cens.amt, each=n.me), n.ec),
        max.effect=rep(max.effect, n.ec*n.ca)))

descr$Eff.bound.choice <- ebc.nms[descr$Eff.bound.choice]

## Now descr contains one row for each combination of the levels of
## the user defined selection variables, `Eff.bound.choice',
## `max.effect' and `cens.amt'. Keep in mind that the names and number
## of these variables is arbitrary. Next we create a skeleton

```

```

## `cpd.PwrGSD' object with a call to the function `cpd.PwrGSD' with
## argument `descr'

test.example.set <- cpd.PwrGSD(descr)

## Now, the newly created object, of class `cpd.PwrGSD', contains
## an element `descr', a component `date', the date created
## and a component `Elements', an empty list of length equal
## to the number of rows in `descr'. Next we do the computation in
## a loop over the rows of `descr'.

n.descr <- nrow(descr)

for(k in 1:n.descr){

  ## First, we copy the original call to the current call,
  ## `Elements[[k]]$call'

  test.example.set$Elements[[k]]$call <- test.example$call

  ## Use the efficacy boundary choice in the kth row of `descr'
  ## to set the efficacy boundary choice in the current call

  test.example.set$Elements[[k]]$call$EfficacyBoundary <-
  parse(text=as.character(descr[k,"Eff.bound.choice"]))[[1]]

  ## Derive the `rhaz' defined by the selection variable "max.effect"
  ## in the kth row of `descr' and use this to set the `rhaz'
  ## components of the current call

  test.example.set$Elements[[k]]$call$rhaz <-
    exp(descr[k,"max.effect"] * log(rhaz))

  ## Derive the censoring components from the selection variable
  ## "cens.amt" in the kth row of `descr' and place that result
  ## into the current call

  test.example.set$Elements[[k]]$call$hc0 <-
  test.example.set$Elements[[k]]$call$hc1 <-
    exp(descr[k, "cens.amt"] * log(hc))

  ## Now the current call corresponds exactly to the selection
  ## variable values in row `k' of `descr'. The computation is
  ## done by calling `update'

  test.example.set$Elements[[k]] <-
    update(test.example.set$Elements[[k]])
  cat(k/n.descr, "\r")
}

## We can create a new `cpd.PwrGSD' object by subsetting on
## the selection variables in `descr':

Elements(test.example.set,
  subset=(substring(Eff.bound.choice, 1,9)=="LanDemets" &
    max.effect >= 1))

```

```
## or we can plot the results -- see the help under `plot.cpd.PwrGSD'

plot(test.example.set, formula = ~ max.effect | stat * cens.amt,
      subset=(substring(Eff.bound.choice, 1,9)=="LanDemets"))

plot(test.example.set, formula = ~ max.effect | stat * cens.amt,
      subset=(substring(Eff.bound.choice, 1,2)=="SC"))

## Notice the appearance of the selection variable `stat' which was
## not defined in the dataset `descr'.

## Recall that each single "PwrGSD" object can contain results
## for a list of test statistics, as in the example shown here where
## we have results on three statistics per component of `Elements'.
## For this reason the variable `stat' can be also be referenced in
## the `subset' or `formula' arguments of calls to this `plot' method,
## and in the `subset' argument of the function `Power' shown below.

## The function `Power' is used to convert the `cpd.PwrGSD' object
## into a dataframe, stacked by rows of `descr' and by `stat'
## (there are three statistics being profiled per each component of
## `Elements'), for generating tables or performing other
## computations.

Power(test.example.set,
      subset=(substring(Eff.bound.choice, 1,2)=="SC" & stat %in% c(1,3)))

## End(Not run)
```

Power

Extract the Power results

Description

Extract the Power results from a compound object into a Stacked Dataframe

Usage

```
Power(object, subset)
```

Arguments

object	an object of class <code>cpd.PwrGSD</code>
subset	you may extract a subset via a logical expression in the variables of the index dataframe, <code>descr</code>

Value

an object of class `cpd.PwrGSD`. See help on that topic for details.

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

See Also

`cpd.PwrGSD` and `PwrGSD`

Examples

```
## See the `cpd.PwrGSD' example
```

Elements	<i>Create a subset of a "cpd.PwrGSD" object</i>
----------	-------------------------------------------------

Description

Create a subset of a `cpd.PwrGSD` object

Usage

```
Elements(object, subset, na.action = na.pass)
```

Arguments

<code>object</code>	an object of class <code>cpd.PwrGSD</code>
<code>subset</code>	you may extract a subset via a logical expression in the variables of the index dataframe, <code>descr</code>
<code>na.action</code>	a method for handling NA values in the variables in the subset expression.

Value

an object of class `cpd.PwrGSD`. See help on that topic for details.

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

See Also

`cpd.PwrGSD` and `PwrGSD`

Examples

```
## See the `cpd.PwrGSD' example
```

plot.cpd.PwrGSD	<i>Plot Method for cpd.PwrGSD objects</i>
-----------------	-------------------------------------------

Description

Creates a trellis plot of type II error probability and power at each interim analysis, stacked, versus an effect size variable, conditioned upon levels of up to two factors.

Usage

```
## S3 method for class 'cpd.PwrGSD':
plot(x, formula, subset, na.action,...)
```

Arguments

x	an object of class <code>cpd.PwrGSD</code>
formula	a one sided formula of the form <code>~ effect f1</code> or <code>~ effect f1 * f2</code> where effect , f1 , and f2 are variables in the indexing dataframe descr , or the special variable stat which may be used when there are multiple test statistics per component of Elements . See the example in the documentation for <code>cpd.PwrGSD</code>
subset	the plot can be applied to a subset of rows of descr via a logical expression on its variables in combination with the special variable, stat when applicable.
na.action	a <code>na.action</code> method for handling NA values
...	other parameters to pass to the R function <code>coplot</code> usually not necessary

Value

Returns the object, **x**, invisibly

Note

This processes the `cpd.PwrGSD` object into a dataframe, stacked on interim looks and then passes the results to the R function `coplot`

Author(s)

Abovementioned `cpd.PwrGSD` processing done by Grant Izmirlian <izmirlian@nih.gov>

References

Chambers, J. M. (1992) *Data for models*. Chapter 3 of *Statistical Models in S* eds J. M. Chambers and T. J. Hastie, Wadsworth & Brooks/Cole.

Cleveland, W. S. (1993) *Visualizing Data*. New Jersey: Summit Press.

See Also

`cpd.PwrGSD Power and Elements`

Examples

```
## See the example in the 'cpd.PwrGSD' documentation
```

GrpSeqBnds

Computes efficacy and futility boundaries

Description

This computes efficacy and futility boundaries for interim analysis and sequential designs. Two sided symmetric efficacy boundaries can be computed by specifying half of the intended total type I error probability in the argument, **Alpha.Efficacy**. Otherwise, especially in the case of efficacy and futility bounds only one sided boundaries are currently computed. The computation allows for two different time scales—one must be the variance ratio, and the second can be a user chosen increasing scale beginning with 0 that takes the value 1 at the conclusion of the trial.

Usage

```
GrpSeqBnds(EfficacyBoundary = LanDemets(alpha = 0.05, spending = ObrienFleming),
           FutilityBoundary = LanDemets(alpha = 0.1, spending = ObrienFleming),
           frac, frac.ii = NULL, drift = rep(0, length(frac)))
```

Arguments

EfficacyBoundary

This specifies the method used to construct the efficacy boundary. The available choices are

- (i) **Lan-Demets(alpha=<total type I error>, spending=<spending function>)**. The Lan-Demets method is based upon a error probability spending approach. The spending function can be set to **ObrienFleming**, **Pocock**, or **Power(rho)**, where **rho** is the the power argument for the power spending function: **rho=3** is roughly equivalent to the O'Brien-Fleming spending function and smaller powers result in a less conservative spending function.
- (ii) **Haybittle(alpha=<total type I error>, b.Haybittle=<user specified boundary point>)**. The Haybittle approach is the simplest, which sets the boundary points equal to **b.Haybittle**, a user specified value (try 3) for all analyses except the last, which is calculated so as to result in the total type I error, set with the argument **alpha**.
- (iii) **SC(alpha=<total type I error>, crit=<threshold for conditional type I error for efficacy stopping>)**. The stochastic curtailment method is based upon the conditional probability of type I error given the current value of the statistic. Under this method, a sequence of boundary points on the standard normal scale (as are boundary points under all other methods) is calculated so that the total probability of type I error, **alpha**, is maintained. This is done by considering the joint probabilities of continuing to the current analysis and then exceeding the threshold at the current analysis. A good value for the threshold value for the conditional type I error, **crit** is 0.90 or greater.
- (iv) User supplied boundary points in the form **c(b1, b2, b3, ..., b_m)**, where **m** is the number of looks.

FutilityBoundary

This specifies the method used to construct the futility boundary. The available choices are

- (i) **Lan-Demets(alpha=<total type II error>, spending=<spending function>).**
The Lan-Demets method is based upon a error probability spending approach. The spending function can be set to **ObrienFleming**, **Pocock**, or **Power(rho)**, where **rho** is the the power argument for the power spending function: **rho=3** is roughly equivalent to the O'Brien-Fleming spending function and smaller powers result in a less conservative spending function.
- (ii) **Haybittle(alpha=<total type I error>, b.Haybittle=<user specified boundary point>).** The Haybittle approach is the simplest, which sets the boundary points equal to **b.Haybittle**, a user specified value (try 3) for all analyses except the last, which is calculated so as to result in the total type II error, set with the argument **alpha**.
- (iii) **SC(alpha=<total type II error>, crit=<threshold for conditional type II error for futility stopping>, drift.end=<projected drift at end of trial>).** The stochastic curtailment method is based upon the conditional probability of type II error given the current value of the statistic. Under this method, a sequence of boundary points on the standard normal scale (as are boundary points under all other methods) is calculated so that the total probability of type II error, **alpha**, is maintained. This is done by considering the joint probabilities of continuing to the current analysis and then exceeding the threshold at the current analysis. A good value for the threshold value for the conditional type I error, **crit** is 0.90 or greater.
- (iv) User supplied boundary points in the form **c(b1, b2, b3,..., b_m)**, where **m** is the number of looks.
- frac** The variance ratio. If the end of trial variance is unknown then normalize all previous variances by the current variance. In this case you must specify a second scale that is monotone increasing from 0 to 1 at the end of the trial. Required.
- frac.ii** The second information scale that is used for type I and type II error probability spending. Optional (see above)
- drift** The drift function of the underlying brownian motion, which is the expected value under the design alternative of the un-normalized weighted log-rank statistic, then normalized to have variance one when the variance ratio equals 1. See the examples below.

Value

An object of class **boundaries** with components: "table" "frac" "frac.ii" "drift" "call"

- call** The call that produced the returned results.
- frac** The vector of variance ratios.
- frac.ii** The vector of information ratios for type I and type II error probability spending, which differs from the above if the user sets the argument **frac.ii** to a second scale as mentioned above.
- drift** The drift vector that is required as an argument when futility boundaries are calculated.
- table** A matrix with components
- frac** The information ratio for type I and type II error probability spending.
- b.f** The calculated futility boundary (if requested).

alpha.f	The type II error probability spent at that analysis (if doing futility bounds).
cum-alpha.f	Cumulative sum of alpha.f (if doing futility bounds).
b.e	The calculated efficacy boundary.
alpha.e	The type I error probability spent at that analysis.
cum-alpha.e	Cumulative sum of alpha.e .

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

References

Gu, M.-G. and Lai, T.-L. "Determination of power and sample size in the design of clinical trials with failure-time endpoints and interim analyses." Controlled Clinical Trials 20 (5): 423-438. 1999

Izmirlian, G. "The PwrGSD package." NCI Div. of Cancer Prevention Technical Report. 2004

Jennison, C. and Turnbull, B.W. (1999) Group Sequential Methods: Applications to Clinical Trials Chapman & Hall/Crc, Boca Raton FL

Proschan, M.A., Lan, K.K.G., Wittes, J.T. (2006), corr 2nd printing (2008) Statistical Monitoring of Clinical Trials A Unified Approach Springer Verlag, New York

See Also

PwrGSD

Examples

```
## NOTE: In an unweighted analysis, the variance ratios and event ratios
## are the same, whereas in a weighted analysis, they are quite different.
##
## For example, in a trial with 7 or so years of accrual and maximum follow-up of 20 years
## using the stopped Fleming-Harrington weights, 'WtFun' = "SFH", with paramaters
## 'ppar' = c(0, 1, 10) we might get the following vector of variance ratios:

frac    <- c(0.006995655, 0.01444565, 0.02682463, 0.04641363, 0.0585665,
             0.07614902, 0.1135391, 0.168252, 0.2336901, 0.3186155, 0.4164776,
             0.5352199, 0.670739, 0.8246061, 1)

## and the following vector of event ratios:

frac.ii <- c(0.1494354, 0.1972965, 0.2625075, 0.3274323, 0.3519184, 0.40231,
             0.4673037, 0.5579035, 0.6080742, 0.6982293, 0.7671917, 0.8195019,
             0.9045182, 0.9515884, 1)

## and the following drift under a given alternative hypothesis

drift <-  c(0.06214444, 0.1061856, 0.1731267, 0.2641265, 0.3105231, 0.3836636,
            0.5117394, 0.6918584, 0.8657705, 1.091984, 1.311094, 1.538582,
            1.818346, 2.081775, 2.345386)

## JUST ONE SIDED EFFICACY BOUNDARY
```

```

## In this call, we calculate a one sided efficacy boundary at each of 15 analyses
## which will occur at the given (known) variance ratios, and we use the variance
## ratio for type I error probability spending, with a total type I error probability
## of 0.05, using the Lan-Demets method with O'Brien-Fleming spending (the default).

gsb.all.just.eff <- GrpSeqBnds(frac=frac,
                             EfficacyBoundary=LanDemets(alpha=0.05, spending=ObrienFleming))

## ONE SIDED EFFICACY AND FUTILITY BOUNDARIES
## In this call, we calculate a one sided efficacy boundary at each of 15 analyses
## which will occur at the given (known) variance ratios, and we use the variance
## ratio for type I and type II error probability spending, with a total type I error
## probability of 0.05 and a total type II error probability of 0.10, using the Lan-Demets
## method with O'Brien-Fleming spending (the default) for both efficacy and futility.

gsb.all.eff.fut <- GrpSeqBnds(frac=frac,
                             EfficacyBoundary=LanDemets(alpha=0.05, spending=ObrienFleming),
                             FutilityBoundary=LanDemets(alpha=0.10, spending=ObrienFleming),
                             drift=drift)

## Now suppose that we are performing the 7th interim analysis. We don't know what the variance
## will be at the end of the trial, so we normalize variances of the current and previous
## statistics by the variance of the current statistic. This is equivalent to the following
## length 7 vector of variance ratios:

frac7 <- frac[1:7]/frac[7]

## To proceed under the "unknown variance at end of trial" case, we must use a second
## scale for spending type I and II error probability. Unlike the above scale
## which is renormalized at each analysis to have value 1 at the current analysis, the
## alpha spending scale must be monotone increasing and attain the value 1 only at the
## end of the trial. A natural choice is the event ratio, which is known in advance if
## the trial is run until a required number of events is obtained, a so called
## maximum information trial:

frac7.ii <- frac.ii[1:7]

## the first seven values of the drift function

drift7 <- drift[1:7]/frac[7]^0.5

gsb.1st7.eff.fut <- GrpSeqBnds(frac=frac7, frac.ii=frac7.ii,
                             EfficacyBoundary=LanDemets(alpha=0.05, spending=ObrienFleming),
                             FutilityBoundary=LanDemets(alpha=0.10, spending=ObrienFleming),
                             drift=drift7)

## Of course there are other options not covered in these examples but this should get you
## started

```

SimGSB

Verifies the results of "GrpSeqBnds" via simulation

Description

Verifies the results of GrpSeqBnds via simulation

Usage

```
SimGSB(object, nsim = 1e+05, ...)
```

Arguments

object an object of class either **boundaries** or **PwrGSD**

nsim number of simulations to do

... if **object** is of class **PwrGSD** and there are more than one statistic under investigation, then you may specify an argument **stat**. The default value is 1, meaning the first one.

Value

A tabulation of the results

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

See Also

GrpSeqBnds

Examples

```
## none as yet
```

as.boundaries	<i>Convert a "PwrGSD" object to a "boundaries" object</i>
----------------------	-----------------------------------------------------------

Description

Convert a **PwrGSD** object to a **boundaries** object

Usage

```
as.boundaries(object, ...)
```

Arguments

object an object of class **PwrGSD**

... if **object** is of class **PwrGSD** and there are more than one statistic under investigation, then you may specify an argument **stat**. The default value is 1, meaning the first one.

Value

an object of class **boundaries**. See the documentation for **GrpSeqBnds**

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

See Also

GrpSeqBnds

Examples

```
## none as yet
```

wtdlogrank	<i>Weighted log-rank test</i>
------------	-------------------------------

Description

Computes a two sample weighted log-rank statistic with events weighted according to one of the available weighting function choices

Usage

```
wtdlogrank(formula = formula(data), data = parent.frame(), WtFun = c("FH", "SFH", "Ramp"),
  param = c(0, 0), sided = c(2, 1), subset, na.action, w = FALSE)
```

Arguments

formula	a formula of the form <code>Surv(Time, Event) ~ arm</code> where <code>arm</code> is a dichotomous variable with values 0 and 1.
data	a dataframe
WtFun	a selection from the available list: “FH” (Fleming-Harrington), “SFH” (stopped Fleming-Harrington) or “Ramp”. See param in the following line.
param	Weight function parameters. Length and interpretation depends upon the selected value of WtFun : If WtFun == <code>"FH"</code> then param is a length 2 vector specifying the power of the pooled (across arms) kaplan meier estimate and its complement. If WtFun == <code>"SFH"</code> then param is a length 3 vector with first two components as in the “FH” case, and third component the time (in the same units as the time to event) at which the “FH” weight function is capped off at its current value. If WtFun == <code>"Ramp"</code> then param is of length 1 specifying the time (same units as time to event) at which events begin to get equal weight. The “Ramp” weight function is a linearly increasing deterministic weight function which is capped off at 1 at the user specified time.
sided	One or Two sided test? Set to 1 or 2
subset	Analysis can be applied to a subset of the dataframe based upon a logical expression in its variables
na.action	Method for handling NA values in the covariate, arm
w	currently no effect

Value

An object of class **survtest** containing components

pn	sample size
wtyp	internal representation of the WtFun argument
par	internal representation of the param argument
time	unique times of events accross all arms
nrisk	number at risk accross all arms at each event time
nrisk1	Number at risk in the experimental arm at each event time
nevent	Number of events accross all arms at each event time
nevent1	Number of events in the experimental arm at each event time
wt	Values of the weight function at each event time
pntimes	Number of event times
stat	The un-normalized weighted log-rank statistic, i.e. the summed weighted observed minus expected differences at each event time
var	Variance estimate for the above
UQt	Cumulative sum of increments in the sum resulting in stat above
varQt	Cumulative sum of increments in the sum resulting in var above
var1t	Cumulative sum of increments in the sum resulting in the variance of an unweighted version of the statistic
pu0	person units of follow-up time in the control arm
pu1	person units of follow-up time in the intervention arm
n0	events in the control arm
n1	events in the intervention arm
n	sample size, same as pn
call	the call that created the object

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

References

Harrington, D. P. and Fleming, T. R. (1982). A class of rank test procedures for censored survival data. *Biometrika* **69**, 553-566.

See Also

IntSurvDiff

Examples

```
library(PwrGSD)
data(lung)
fit.wlr <- wtdlogrank(Surv(time, I(status==2))~I(sex==2), data=lung, WtFun="SFH", param=c(0,1,300))
```

IntSurvDiff*Weighted Integrated Survival function test*

Description

Computes a two sample weighted integrated survival function log-rank statistic with events weighted according to one of the available weighting function choices

Usage

```
IntSurvDiff(formula = formula(data), data = parent.frame(), WtFun = c("FH", "SFH", "Ramp"),
            param = c(0, 0), sided = c(2, 1), subset, na.action, w = FALSE)
```

Arguments

formula	a formula of the form <code>Surv(Time, Event) ~ arm</code> where arm is a dichotomous variable with values 0 and 1.
data	a dataframe
WtFun	a selection from the available list: “FH” (Fleming-Harrington), “SFH” (stopped Fleming-Harrington) or “Ramp”. See param in the following line.
param	Weight function parameters. Length and interpretation depends upon the selected value of WtFun : If WtFun == <code>"FH"</code> then param is a length 2 vector specifying the power of the pooled (across arms) kaplan meier estimate and its complement. If WtFun == <code>"SFH"</code> then param is a length 3 vector with first two components as in the “FH” case, and third component the time (in the same units as the time to event) at which the “FH” weight function is capped off at its current value. If WtFun == <code>"Ramp"</code> then param is of length 1 specifying the time (same units as time to event) at which events begin to get equal weight. The “Ramp” weight function is a linearly increasing deterministic weight function which is capped off at 1 at the user specified time.
sided	One or Two sided test? Set to 1 or 2
subset	Analysis can be applied to a subset of the dataframe based upon a logical expression in its variables
na.action	Method for handling NA values in the covariate, arm
w	currently no effect

Value

An object of class **survtest** containing components

pn	sample size
wtyp	internal representation of the WtFun argument
par	internal representation of the param argument
time	unique times of events accross all arms
nrisk	number at risk accross all arms at each event time
nrisk1	Number at risk in the experimental arm at each event time

<code>nevent</code>	Number of events accross all arms at each event time
<code>nevent1</code>	Number of events in the experimental arm at each event time
<code>wt</code>	Values of the weight function at each event time
<code>pntimes</code>	Number of event times
<code>stat</code>	The un-normalized weighted log-rank statistic, i.e. the summed weighted observed minus expected differences at each event time
<code>var</code>	Variance estimate for the above
<code>pu0</code>	person units of follow-up time in the control arm
<code>pu1</code>	person units of follow-up time in the intervention arm
<code>n0</code>	events in the control arm
<code>n1</code>	events in the intervention arm
<code>n</code>	sample size, same as <code>pn</code>
<code>call</code>	the call that created the object

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

References

Weiland S, Gail MH, James BR, James KL. (1989). A family of nonparametric statistics for comparing diagnostic makers with paired or unpaired data. *Biometrika* **76**, 585-592.

See Also

`wtdlogrank`

Examples

```
library(PwrGSD)
data(lung)
fit.isd <- IntSurvDiff(Surv(time, I(status==2))~I(sex==2), data=lung, WtFun="SFH", param=c(0,1,300))
```

mysurvfit

My Survfit

Description

Computes numbers at risk, numbers of events at each unique event time within levels of a blocking factor

Usage

```
mysurvfit(formula = formula(data), data = parent.frame(), subset, na.action = na.fail)
```


Arguments

<code>formula</code>	Should be a formula of the form <code>Surv(ti, ev) ~ block</code> where <code>block</code> is the blocking factor. It need not be a factor per se but should have relatively few discrete levels. Sorry, no staggered entry allowed at present
<code>data</code>	a dataframe
<code>subset</code>	you can subset the analysis via logical expression in variables in the dataframe
<code>na.action</code>	pass a method for handling NA values in <code>block</code> such as <code>na.omit</code> , or <code>na.fail</code>

Value

A dataframe of $2 \times \text{NLEV} + 1$ columns where NLEV is the number of levels of the factor `block`.

<code>time</code>	The sorted vector of unique event times from all blocks
<code>nrisk1</code>	The number at risk in block level 1 at each event time
<code>nevent1</code>	The number of events in block level 1 at each event time
<code>...</code>	
<code>nriskNLEV</code>	The number at risk in block level NLEV at each event time
<code>neventNLEV</code>	The number of events in block level NLEV at each event time

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

Examples

```
library(PwrGSD)
data(lung)

fit.msf <- mysurvfit(Surv(time, I(status==2)) ~ sex, data=lung)

fit.msf
## Not run:
plot(fit.msf)

## End(Not run)
```

agghaz

Aggregated Hazard

Description

Computes the MLE for the model that assumes piecewise constant hazards on intervals defined by a grid of points. One applications for example is to calculate monthly hazard rates given numbers of events, numbers at risk and event times reported to the day. Can also handle time to event data stratified on a blocking factor.

Usage

```
agghaz(t.agg, time, nrisk, nevent)
```

Arguments

<code>t.agg</code>	Vector defining intervals upon which the user wants constant hazard rates.
<code>time</code>	Event times, possibly stratified on a blocking factor into multiple columns, in units that occur in enough numbers per interval specified above. If there is just a single column then it must be in column form (see example below).
<code>nrisk</code>	Numbers at risk at specified event times
<code>nevent</code>	Numbers of events at specified event times

Value

<code>time.a</code>	User supplied left-hand endpoints of intervals of hazard constancy
<code>nrisk.a</code>	Numbers at risk on specified intervals
<code>nevent.a</code>	Numbers of events on specified intervals

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

Examples

```
library(PwrGSD)
data(lung)
fit.msf <- mysurvfit(Surv(time, I(status==2)) ~ sex, data=lung)

## A single stratum:
with(fit.msf$Table, agghaz(30, time, cbind(nrisk1), cbind(nevent1)))

## Multiple strata--pooled and group 1:
with(fit.msf$Table, agghaz(30, time, cbind(nrisk1+nrisk2,nrisk1), cbind(nevent1+nevent2,nevent1)))
```

mystack	<i>Stack a dataset</i>
---------	------------------------

Description

Given a dataframe containing one or more variables named with a common prefix, this function creates a stacked dataset with one set of observed values of the variables (in order of occurrence) per line.

Usage

```
mystack(object, fu.vars, create.idvar = FALSE)
```

Arguments

<code>object</code>	a dataframe containing one or more variables named with a common prefix
<code>fu.vars</code>	a list of the unique prefixes
<code>create.idvar</code>	Do you want to add an ID variable with a common value given to all records resulting from a given input record? Default is FALSE

Value

A stacked dataframe

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

Examples

```
## none as yet
```

CDFOR2LRR

Convert CDF Odds Ratio to Logged Relative Risks

Description

A concise (1-5 lines) description of what the function does.

Usage

```
CDFOR2LRR(tcut, tmax, h0, CDFOR)
```

Arguments

tcut	Describe tcut here
tmax	Describe tmax here
h0	Describe h0 here
CDFOR	Describe CDFOR here

Details

If necessary, more details than the description above

Value

Describe the value returned If it is a LIST, use

comp1	Description of 'comp1'
comp2	Description of 'comp2'
...	

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

See Also

objects to See Also as `help`,

Examples

```
## none as yet
```

CY2T0Shaz

*Calender year rates to Study Year Rates***Description**

Given the cutpoints at which the hazard is to be constant, the values taken by the calender year rates and the calender time offset from the start of the trial at which randomization ended, this function converts to time on study rates, assuming uniform accrual.

Usage

```
CY2T0Shaz(tcut, t.eor, m, verbose = FALSE)
```

Arguments

<code>tcut</code>	Left hand endpoints of intervals on which time on study hazard is taken to be constant
<code>t.eor</code>	Time offsest from the beginning of the trial at which randomization ended
<code>m</code>	Annual calender time rates
<code>verbose</code>	do you want to see alot of debugging info—defaults to FALSE

Value

```
hazard = h, table = attr(obj., "tbl")
```

<code>hazard</code>	time on study hazard values taken on intervals specified by the argument <code>tcut</code>
---------------------	--------------------------------------------------------------------------------------------

<code>table</code>	a table containg the observed and fitted values
--------------------	-------------------------------------------------

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

Examples

```
## none as yet
```

CRRtoRR

*Cumulative-risk ratios to risk ratios***Description**

Given a vector of cumulative-risk ratios, computes risk ratios

Usage

```
CRRtoRR(CRR, DT, h = NULL)
```

Arguments

CRR	vector of cumulative risk ratios of length m
DT	vector of time increments upon which the cumulative ratios represent. For example if the hazard takes values h_1, h_2, \dots, h_m on the intervals $[t_1, t_2), [t_2, t_3), \dots, [t_m, t_{m+1})$ then DT will be $c(t_2 - t_1, t_3 - t_2, \dots, t_{m+1} - t_m)$
h	The hazard in the reference arm, of length m

Value

The vector of risk ratios at the **m** time points

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

Examples

```
## none as yet
```

RCM2RR

Relative cumulative mortality to Relative Risk

Description

Relative cumulative mortality to Relative Risk

Usage

```
RCM2RR(tlook, tcut.i, h.i, h0th, accru, rcm)
```

Arguments

```
tlook
tcut.i
h.i
h0th
accru
rcm
```

Value

Describe the value returned If it is a **LIST**, use

```
comp1      Description of 'comp1'
comp2      Description of 'comp2'
```

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

See Also

objects to See Also as **help**,

Examples

```
## none as yet
```

RR2RCM	<i>Relative risk to Relative Cumulative Mortality</i>
--------	-------------------------------------------------------

Description

Relative risk to Relative Cumulative Mortality

Usage

```
RR2RCM(tlook, tcut.i, tcut.ii, h, rr, h0th, accru)
```

Arguments

tlook	Describe tlook here
tcut.i	Describe tcut.i here
tcut.ii	Describe tcut.ii here
h	Describe h here
rr	Describe rr here
h0th	Describe h0th here
accru	Describe accru here

Value

Describe the value returned If it is a LIST, use

comp1	Description of 'comp1'
comp2	Description of 'comp2'

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

See Also

objects to See Also as **help**,

Examples

```
## none as yet
```

lookup	<i>Lookup values for a piecewise constant function</i>
--------	--------------------------------------------------------

Description

Given the values and lefthand endpoints for intervals of constancy, lookup values of the function at arbitrary values of the independent variable.

Usage

```
lookup(xgrid, ygrid, x, y0 = 0)
```

Arguments

xgrid	Lefthand endpoints of intervals of constancy
ygrid	Values on these intervals, of same length as xgrid
x	Input vector of arbitrary independent variables.
y0	Value to be returned for values of x that are smaller than <code>min(xgrid)</code> .

Value

Describe the value returned If it is a LIST, use

comp1	Description of 'comp1'
comp2	Description of 'comp2'

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

Examples

```
## none as yet
```

lung	<i>Mayo Clinic Lung Cancer Data</i>
------	-------------------------------------

Description

Survival in patients with lung cancer at Mayo Clinic. Performance scores rate how well the patient can perform usual daily activities.

Usage

```
data(lung)
```

Format

inst:	Institution code
time:	Survival time in days
status:	censoring status 1=censored, 2=dead
age:	Age in years
sex:	Male=1 Female=2
ph.ecog:	ECOG performance score (0=good 5=dead)
ph.karno:	Karnofsky performance score (bad=0-good=100) rated by physician
pat.karno:	Karnofsky performance score rated by patient
meal.cal:	Calories consumed at meals
wt.loss:	Weight loss in last six months

Source

Terry Therneau

Surv

Create a Survival Object

Description

Create a survival object, usually used as a response variable in a model formula.

Usage

```
Surv(time, event)
or
Surv(time, time2, event, type=, origin=0)
```

Arguments

time	for right censored data, this is the follow up time. For interval data, the first argument is the starting time for the interval.
event	The status indicator, normally 0=alive, 1=dead. Other choices are T/F (TRUE = death) or 1/2 (2=death). For interval censored data, the status indicator is 0=right censored, 1= event at time , 2=left censored, 3=interval censored. Although unusual, the event indicator can be omitted, in which case all subjects are assumed to have an event.
time2	ending time of the interval for interval censored or counting process data only. Intervals are assumed to be open on the left and closed on the right, (start , end]. For counting process data, event indicates whether an event occurred at the end of the interval.
type	character string specifying the type of censoring. Possible values are "right", "left", "counting", "interval", or "interval2". The default is "right" or "counting" depending on whether the time2 argument is absent or present, respectively.

origin for counting process data, the hazard function origin. This is most often used in conjunction with a model containing time dependent strata in order to align the subjects properly when they cross over from one strata to another.

Value

An object of class **Surv**. There are methods for **print**, **is.na**, and subscripting survival objects. To include a survival object inside a data frame, use the **I()** function. **Surv** objects are implemented as a matrix of 2 or 3 columns.

In the case of **is.Surv**, a logical value T if **x** inherits from class "**Surv**", otherwise an F.

DETAILS

In theory it is possible to represent interval censored data without a third column containing the explicit status. Exact, right censored, left censored and interval censored observation would be represented as intervals of (a,a), (a, infinity), (-infinity,b), and (a,b) respectively; each specifying the interval within which the event is known to have occurred.

If **type = "interval2"** then the representation given above is assumed, with NA taking the place of infinity. If **type="interval"** **event** must be given. If **event** is 0, 1, or 2, the relevant information is assumed to be contained in **time**, the value in **time2** is ignored, and the second column of the result will contain a placeholder.

Presently, the only methods allowing interval censored data are the parametric models computed by **survreg**, so the distinction between open and closed intervals is unimportant. The distinction is important for counting process data and the Cox model.

The function tries to distinguish between the use of 0/1 and 1/2 coding for left and right censored data using **if (max(status)==2)**. If 1/2 coding is used and all the subjects are censored, it will guess wrong. Use 0/1 coding in this case.

See Also

wtdlogrank, **IntSurvDiff**, **mysurvfit**.

Examples

```
## none as yet
```

DX	<i>function to do ...</i>
----	---------------------------

Description

A concise (1-5 lines) description of what the function does.

Usage

```
DX(x)
```

Arguments

x Describe **x** here

Details

If necessary, more details than the description above

Value

Describe the value returned If it is a LIST, use

comp1 Description of 'comp1'

comp2 Description of 'comp2'

...

Author(s)

Grant Izmirlian <izmirlian@nih.gov>

References

put references to the literature/web site here

Examples

```
## none as yet
```

paste

The paste operator

Description

A binary operator shortcut for paste(x,y)

Usage

```
x %,% y
```

Arguments

x a character string

y a character string

Value

```
paste(x, y, sep="")
```

Author(s)

Grant Izmirlian (izmirlian@nih.gov)

Examples

```
library(PwrGSD)
"var" %,% (1:10)
```

Index

- *Topic **character**
 - paste, 34
- *Topic **datasets**
 - lung, 31
- *Topic **data**
 - cpd.PwrGSD, 9
 - Elements, 14
 - mystack, 26
 - Power, 13
- *Topic **design**
 - as.boundaries, 20
 - GrpSeqBnds, 15
 - PwrGSD, 1
- *Topic **hplot**
 - plot.cpd.PwrGSD, 14
- *Topic **htest**
 - as.boundaries, 20
 - GrpSeqBnds, 15
 - PwrGSD, 1
- *Topic **manip**
 - agghaz, 25
- *Topic **misc**
 - DX, 33
 - lookup, 31
 - SimGSB, 19
- *Topic **survival**
 - agghaz, 25
 - CDFOR2LRR, 27
 - CRRtoRR, 28
 - CY2TOShaz, 28
 - IntSurvDiff, 22
 - mysurvfit, 24
 - plot.cpd.PwrGSD, 14
 - PwrGSD, 1
 - RCM2RR, 29
 - RR2RCM, 30
 - Surv, 32
 - wtdlogrank, 21
- %,% (*paste*), 34
- agghaz, 25
- as.boundaries, 20
- CDFOR2LRR, 27
- cpd.PwrGSD, 8, 9, 13–15
- CRRtoRR, 28
- CY2TOShaz, 28
- DX, 33
- Elements, 10, 14, 15
- GrpSeqBnds, 15, 20
- help, 27, 30
- IntSurvDiff, 22, 22, 33
- lookup, 31
- lung, 31
- mystack, 26
- mysurvfit, 24, 33
- paste, 34
- plot.cpd.PwrGSD, 10, 14
- Power, 10, 13, 15
- PwrGSD, 1, 13, 14, 18
- RCM2RR, 29
- RR2RCM, 30
- SimGSB, 19
- Surv, 32
- wtdlogrank, 21, 24, 33